





JUNE 29, 2023 | DR. LAURA LLERAS FORERO AND INES KRISTINA HARTMANN

Good Cell Culture Practice

How to Improve the Reproducibility of Your Experiments

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Your Experts Today



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Agenda

Introduction

- Cell Passage, Confluency, and Cell Count
- Cell Seeding
- Cell Incubation
- Conclusions and F
 - **Conclusions and Recommendations**



What Is Reproducibility?

- 1. The ability to replicate the results from a published study using the same methods and materials.
- 2. Are the methodology and results presented in sufficient detail to enable replication?



Reproducibility in Science

Is there a reproducibility crisis?*



Have you ever established procedures for reproducibility?*



*1,576 researchers surveyed

Cultured Cells as Model Systems

Benefits:

- > Cells retain many functions and properties of their parental tissue
- > Cells serve as a model system to test new therapeutic approaches
- > Renewable resource of cell material

Risks:

- > Contamination with microorganisms
- > Cross-contamination of different cell lines
- > Genetic drift during cultivation
- > Response to different cultivation conditions



"Cell culture sometimes feels like a black art, with everyone having their own preferred method"

- Dr. Delcassian, MIT (Mass., USA). Technology Networks Cell Science 2018

Cell Passage, Confluency, and Cell Count

02

The Human Eye Is Not Quantitative







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Irregular Cell Culture Process



Optimal Timepoint for Passaging

Optimizing cell growth

- > Plot a cell growth curve
- > Don't let the cultures overgrow
- > Check cell viability
- > Do not use cultures with <80% viability





^a Centre for Cell Engineering Josenh Black Building University of Glasgow G12.800 Glasgow UK

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Cell Passage Influences Cell Culture Reproducibility



Cell Morphology Changes with Different Passages



Circularity (a.u.)

Osaki, T et al. Sci Rep 7, 1897 (2017).

Cell Morphology Changes with Different Passages



Li et al. Cell Tissue Res (2007).

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Avoid Prolonged Passaging

- > Use high-quality cells as a starting point
- > Establish a reference cell stock
- > Determine the safe passage number by establishing baselines
 - > Expression/presence of proteins of interest
 - > Proliferation rates
- > Monitor morphology and document culture maintenance
- > Stop active cultivation of cells that are not used

Cell Count Is Essential for Experimental Reproducibility



Niepel et al. Cell Systems (2019)

Cell Count Affects the End Experimental Result



Niepel et al. Cell Systems (2019)



Introduction



Cell Culture Workflow

Routine Maintenance



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Prepare Your Workspace

If you handle different cell lines:

- > Work with one cell line at a time
- > Disinfect the workspace in between
- > Dedicate aliquots/media bottles for each cell line







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eppendorf Know Your Cells' Growth Behavior and Morphology





Check for Contamination



Bacteria

Fungus

Yeast





Cell cultures should be tested for mycoplasma contamination regularly.



Seeding Cells

Filling a plate is time consuming.

- > Cell sedimentation in the tube is quite fast
- The longer the seeding process, the higher the decrease in cell number
- > Make sure to resuspend in between

The formation of air bubbles.

- > Culture medium tends to foam
- > Air bubbles can hinder cell attachment
- > Avoid harsh and fast pipetting





Influence of Pipetting Systems

Air-cushion

- > Variable influencing factors
- Optimal for aqueous solutions
- > Prone to contamination by aerosols (-> filter tips)



Tip with integrated piston No air above the liquid

Positive displacement

- > Unaffected by physical properties of liquid
- > Suitable for "problematic" liquids
- > Contamination free

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Pipetting Different Liquids

Common dissolvents can be problematic

- > Ethanol (volatile liquid)
- > DMSO (viscous liquid)



Air-cushion vs. positive displacement

Good Pipetting Practice (Air-Cushion)

- Choose the smallest pipette (for 100 μL, a 10–100 μL is prefered rather than a 100–1000 μL)
- > Hold the pipette vertical when aspirating liquid
- > Only slightly immerse the tip in the liquid
- > Keep a constant immersion depth
- > While dispensing, keep a constant angle of 45°



Liquid pipetted with a varying angle during dispensing







Mixing and Pipetting Technique

Pipetting can affect assay results.



Source: InCelligence



How to Achieve Homogeneous Cell Adhesion





Density-Dependent Cell Behavior





Low density



Medium density



High density





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Constant Documentation



Clear and traceable cell identification



Culture conditions and procedures



Reference images



Everyday culture practice

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| HeLa seeding density | | | | | | | | |
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| Date | Person | Passage Nr. | cells/mL | Viability | Split Ratio | Comments | | | | | | 1 | |
| 16.04.05 | Julia | 16 | - | | | Cells seeding intitiated | | | | | | | |
| 16.04.06 | Julia | 17 | | | | Medium changed | | | | | | | |
| 16.04.07 | Gabriela | 20 | 1 x 10^6 | 45% | 1/3 | | | | | | | | |
| 16.04.08 | Julia | 16 | 3 x 10^4 | 43% | 1/5 | | | | | | | | |
| 16.04.09 | Julia | 18 | - | | | | | ÷ | | | | - | |
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Increase reproducibility by minimizing user-dependent variations

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Introduction



Cell Culture Workflow



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Provide Stable Incubation Conditions

Reduce traffic in and out:

- > Incubators with segmented inner doors
- > Recovery of temperature and CO₂ (O₂) level
- > No set point overshoot
- > Separate incubators for maintenance and experiment, etc.









Avoid Vibrations

Incubator door openings



Closed door



Frequent door openings



Moving culture vessels after seeding



Not moving



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Manual moving

Other vibration sources in the lab:

- > Centrifuges, freezers, air conditioners
- > CO_2 incubators with a fan



Minimize Disturbances—Stay Organized

Organize to reduce the number and duration of incubator door openings.



- > Optimal incubator content organization depends on lab routines
- > Segmented inner doors decrease recovery times



Avoid Contamination Risks

Clean and decontaminate on a regular basis.

- > Use incubators with a seamless chamber
- > Disassemble and clean inner parts
- > Clean/disinfect inner walls
- > Run high disinfection routine (optional)





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Monitor Incubation Parameters

Reproducible culture conditions

- > Check for door openings during incubation
- > Download performance data for documentation









Conclusions and Recommendations

- 1. To increase reproducibility in your cell count and confluency measurements, include an impartial measurement method not based on personal estimation
- 2. Track population doubling times and profile growth curve characteristics periodically
- 3. When receiving a new batch of cells, test for mycoplasma, authenticate the cells, and make a frozen aliquot
- 4. Establish quality control protocols in your laboratory
- 5. Properly describe everything in the methodology and perform statistical analysis



Conclusions and Recommendations



Reproducible experiments and culture conditions:

- > For seeding cells, choose the proper pipette and pipetting technique
- > For experiments, seed the cells from a pre-diluted master mix and do not let the cells sediment
- Apply continuous proper documentation that can be shared (e.g., with an electronic lab notebook)
- > Avoid vibrations and minimize disturbances in your incubation environment (fan-less design, organized incubators, split doors)
- > Check the data log on your device to track device performance and download it if needed





Thank you for your attention

The CM30 Incubation Monitoring Systems





Product solutions



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- ✓ Saves up to 8,300€/\$9,000 over five years
- ✓ Up to 25% more usable space

Eppendorf.com/co2-incubators





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